

- 4 Schafer P, Braun R W, Mohring K, et al. Quantitative determination of human cytomegalovirus target sequences in peripheral blood leukocytes by nested polymerase chain reaction and temperature gradient gel electrophoresis. *J Gen Virol*, 1993, **74** (12): 2699~ 2707
- 5 Kappes S, Milde Langosch K, Kressin P, et al. p53 mutations in ovarian tumors, detected by temperature gradient gel electrophoresis, direct sequencing and immunohistochemistry. *Int J Cancer*, 1995, **64** (1): 52~ 59
- 6 Schlechte H H, Schnorr D, Loning T, et al. Mutation of the tumor suppressor gene p53 in human prostate and bladder cancers—investigation by temperature gradient gel electrophoresis (TGGE). *J Urol*, 1997, **157** (3): 1049~ 1053
- 7 Kneba M, Bolz I, Linke B, et al. Characterization of clone-specific rearrangement T-cell receptor gamma chain genes in lymphomas and leukemias by the polymerase chain reaction and DNA sequencing. *Blood*, 1994, **84** (2): 574~ 581
- 8 Horn D, Robinson P N, Boddrich A, et al. Three novel mutations of the NF1 gene detected by temperature gradient gel electrophoresis of exons 5 and 8. *Electrophoresis*, 1996, **17** (10): 1559~ 1563

**The Technique of Temperature Gradient Gel Electrophoresis and Its Applications.** CHEN Han-

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**Abstract** Temperature gradient gel electrophoresis (TGGE) is a new and powerful electrophoresis method for separation of nucleic acids (DNA and RNA) and analysis of sequence variations. The TGGE method uses the melting temperature ( $T_m$ ) as an important parameter to identify DNA, which differs in sequence among a mixture of molecules of the same size but different conformation. With the advantages of high-resolution capability, high reproducibility and timesaving process, the TGGE technique has been applied broadly in the field of molecular biology.

**Key words** temperature gradient gel electrophoresis, melting temperature, sequence variation, point mutation

## 一种改良的从银染的聚丙烯酰胺凝胶中回收 DNA 的方法

*Clin Chem*, 1998, **44** (9) 报道了 Steven 等介绍的从聚丙烯酰胺凝胶上将 DNA 转移到 DEAE 膜上的技术。此技术能同时进行多个样品的处理和纯化，被纯化的差异显示法 (DDA) 产物电泳条带单一，分离的范围为 100~ 700 bp 的 DNA，回收的 DNA 足够作数次重扩增反应，且重新扩增的条件不变，重复性高。此技术不需要特殊仪器和试剂，操作简便在 DDA 中应用十分广泛。

细胞遗传学正常的羊水细胞在二倍的常氏 (CISM) 细胞培养液中生长。信使 RNA 用寡聚 dT/链霉抗生物素蛋白磁珠吸附系统分离，以 MMLV 逆转录酶、寡聚 dT 为引物 (5'-TTTTTTTTTTGC)，将其转变成 cDNA。DDA 分析，用 20 μl 反应液。1×PCR 缓冲液，含 1.5 mmol/L MgCl<sub>2</sub>，20 μmol/L dNTP，0.5 μmol/L 寡聚 dT 引物，0.25 μmol/L 上游引物 (5'-CTGCTTGTATG, 5'-GATCCAGTC, 5'-GATCGCATTG, 和 5'-AAACTCCGTC)，30 ng cDNA 和 0.625 U Taq 酶，40 个热循环，95 °C 30 s, 40 °C 1 min, 72 °C 1 min。DNA 产物用乙醇沉淀，再水溶解，用 5% 非变性聚丙烯酰胺凝胶电泳分离，银染显色。

含 DDA 扩增产物的聚丙烯酰胺干胶用小刀切下，放入 2% 琼脂糖凝胶中近阴极端的孔内，压平，小心地用溶化的较凉的 2% 琼脂糖胶填满。将 DEAE 膜裁成 0.5 cm × 0.75 cm 的条，插入离孔 0.5 cm 处，每孔后插一条，可用小刀帮助插入干 DEAE 条。DNA 以 10 V/cm 电泳 90 min 后，将 DEAE 膜取出，放入小离心管内，用 500 μl 低盐 NET (0.15 mmol/L NaCl, 0.1 mmol/L EDTA, 20 mmol/L Tris, pH 8.0) 缓冲液浸泡，然后用 50 μl 高盐 NET 缓冲液 (1 mol/L NaCl, 0.1 mmol/L EDTA, 20 mmol/L Tris, pH 8.0)，在 65 °C 温育 30 min 以洗脱 DNA。DNA 用 2.5 倍的乙醇沉淀，然后用 15 μl 水溶解。用 5 μl 洗脱 DNA，以上述条件进行扩增，产物连接上 T-载体 (Promega) 并测序。

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