



脂代谢紊乱在肾脏衰老和肾纤维化中的作用*

吴胜泉^{1,2)} 杨萌^{1,2)} 刘新光^{1,2)**}

⁽¹⁾ 广东医科大学衰老研究所, 广东省医学分子诊断重点实验室, 东莞 523808;

⁽²⁾ 广东医科大学生物化学与分子生物学研究所, 东莞 523808)

摘要 肾间质纤维化是终末期肾脏病的病理基础, 肾脏衰老是肾间质纤维化的危险因素。越来越多的研究证明, 脂代谢紊乱会导致肾脏衰老和肾间质纤维化。脂代谢紊乱引起的脂质堆积, 会造成脂毒性和细胞应激性损伤, 从而诱发衰老与细胞外基质 (extracellular matrix, ECM) 的分泌。维持脂代谢稳态有助于减轻肾脏衰老与肾间质纤维化的发生发展。脂代谢途径的关键酶和调控蛋白有望成为改善肾脏衰老和肾间质纤维化的潜在靶点。本综述概括了脂代谢紊乱在肾脏衰老和间质纤维化中的作用, 并对脂代谢中肾脏衰老和间质纤维化的预防靶点和策略进行了总结, 为治疗肾纤维化发现新靶点提供了参考。

关键词 肾脏衰老, 肾间质纤维化, 脂代谢, 肾小管上皮细胞

中图分类号 R365, R692

DOI: 10.16476/j.pibb.2023.0305

衰老是指生物体在其生命过程中结构的退行性改变、功能储备的逐渐丧失、对外部或内部压力的适应能力显著下降, 常常伴随着疾病发生, 甚至有死亡的风险^[1]。在过去几十年中, 由于生活条件、社会经济状况和医疗保健的改善, 人类的预期寿命显著增加, 人口的快速老龄化将是全球的必然趋势。目前, 65岁及以上的人口约占世界人口总数的8.5%, 预计2050年达到17%^[2]。中国65岁及以上的人口预计2050年将增加到4亿, 占人口总数的26.9%, 80岁或以上人口数量将达到1.5亿^[3]。随着人口老龄化进程的加快, 各类衰老相关疾病的发生率也逐渐升高, 给社会带来了沉重的经济和医疗负担。据估算, 慢性肾脏病 (chronic kidney disease, CKD) 会影响全球10%~14%的人口^[4], 而在60岁以上人群中的发病率则高达25%^[5]。年龄增长造成的肾脏衰老和肾功能储备受损, 增加了个体对CKD的易感性^[6]。肾间质纤维化被认为是CKD发展至终末期的主要病理基础, CKD的高发为肾纤维化的防治带来很大的挑战。脂质代谢紊乱是指体内血液及其他组织器官中脂质及其代谢产物和量的异常。脂质代谢紊乱与肾脏疾病密切相关, 它既是许多原发或继发性肾脏病的常见临床表现, 同时也参与了肾脏病的进展。随着肾脏能量代

谢的研究越来越多, 人们对脂代谢紊乱在肾脏疾病发生发展过程中扮演的角色有了新的认识, 本文针对这些研究进展进行综述, 阐述肾脏脂代谢异常产生的原因、脂质累积对细胞造成的影响, 梳理脂代谢异常在肾脏衰老和肾间质纤维化中的作用, 并加深对其分子机制和生物学意义的理解。

1 肾脏衰老与肾间质纤维化

1.1 肾脏衰老

随着年龄的增长, 个体的各类器官发生退行性病变。肾脏特别容易受到年龄相关变化的影响^[7], 被认为是正常衰老过程中变化最显著的器官之一^[8]。肾脏衰老的特征是肾皮质的减少和肾功能受损, 组织学的改变包括肾小球硬化、肾小管萎缩、肾纤维化及肾血管硬化^[9], 功能上的变化主要表现为肾小球滤过率 (glomerular filtration rate, eGFR) 的下降。慢性炎症、氧化应激和肾素-血管紧张素-醛固酮 (renin-angiotensin-aldosterone

* 国家自然科学基金 (81971329, 82200821) 和广东省基础与应用基础研究基金 (2023A1515012838) 资助项目。

** 通讯联系人。

Tel: 0769-22896026, E-mail: liuxg@gdmu.edu.cn

收稿日期: 2023-08-03, 接受日期: 2023-10-26

system, RAAS) 系统活性改变造成的细胞损伤和修复能力下降是诱发肾脏衰老的重要驱动因素^[10]。细胞衰老的常见分子变化在衰老的肾脏中也被鉴定出来, 包括衰老相关- β -半乳糖苷酶 (SA- β -gal) 活性增加、细胞周期抑制蛋白上调、衰老分泌相关表型 (senescence associated secretory phenotype, SASP) 的增多和端粒缩短。肾脏的衰老会引发高血压、胰岛素抵抗等一系列问题^[11], 还会增加个体对急性肾损伤和慢性肾病的易感, 并在肾纤维化的发生和发展过程中发挥关键作用。衰老的肾小管上皮细胞 (tubular epithelial cells, TECs) 是区分患病肾脏和正常肾脏的关键^[12-13]。最近研究表明, 不断清除衰老细胞可减轻与年龄相关的肾功能减退和肾小球硬化^[14]。

1.2 肾间质纤维化

肾间质纤维化是所有慢性和进行性肾病的共同病理特征, 也是导致肾功能衰竭的主要决定因素^[15]。它的定义是细胞外基质 (extracellular matrix, ECM) 在小管和管周毛细血管之间的过度沉积^[16], 主要表现为致纤维化细胞因子分泌增多和ECM的胶原成分聚集, 伴有肾小管萎缩和扩张变形、肾单位破坏及肾小球过滤能力减弱^[4, 17]。在轻度损伤的情况下, 纤维化基质沉积有助于组织修复, 其会在组织修复过程中被吸收^[15]。然而, 在CKD的后期, 纤维化基质沉积过多, 降低了组织修复的能力, 则会破坏器官结构, 减少血液供应, 扰乱器官功能, 最终导致肾衰竭^[15]。CKD过程中肾脏的持续损伤通过不同的信号通路激活肌成纤维细胞, 激活的肌成纤维细胞很大一部分是由TECs转化而来的。TECs脱离细胞接触, 失去上皮特征, 获得成纤维细胞表型, 也可产生ECM, 促进纤维化过程^[18]。转化生长因子 β 1 (transforming growth factor beta-1, TGF- β 1) 被认为是这一过程的主要调节因子^[19]。由于肾纤维化的发病机制较为复杂^[20], 阐明肾脏发生纤维化的分子调控机制, 对于肾纤维化的防治具有重要的意义。

1.3 肾脏衰老与肾间质纤维化的关系

肾间质纤维化是肾脏衰老的微观表现, 衰老的肾脏细胞也可在肾间质纤维化中发挥关键作用^[21]。TECs衰老伴随着SASP的表达上调, 引起促纤维化细胞因子分泌增加, 进而推动肾间质纤维化的发生发展^[22]。TECs中纤维化标志物 α -SMA的蛋白质水平与SA- β -gal的活性以及衰老相关的p16和p21的表达呈正相关^[23-24]。缺血再灌注6周的老年小鼠

有广泛的肾纤维化和白细胞浸润, 这与肾小管中p53表达及SA- β -gal阳性细胞的增加有关^[25]。急性肾损伤诱导近端肾小管上皮细胞衰老产生的促纤维化生长因子, 能够刺激成纤维细胞增殖和胶原的形成^[26]。在人肾小管上皮细胞系HK-2和TECs中, Wnt9a能够上调p16、p19、p53和p21的表达, 促进TGF- β 1的分泌, 进而加快肾成纤维细胞的增殖和活化^[27]。以上数据揭示了肾脏衰老和肾纤维化之间的内在联系, 提示延缓肾脏衰老可能是预防肾间质纤维化的有效途径。

2 肾脏脂代谢紊乱的诱发因素

在正常情况下, 体内的脂质会被储存于脂肪组织中^[28]。然而过量的脂质也会在其他组织中沉积, 其中就包括肾脏^[29]。肾脏脂质代谢失调主要源于3条途径的破坏: 循环中脂肪酸 (fatty acid, FA) 摄取、脂质合成和分解。一旦脂质的摄取/合成超出了与分解的平衡, 肾脏脂质就会累积。因此, 肾脏脂代谢紊乱常见的原因主要是血脂异常、脂质合成过多和脂质分解的减少。

考虑到大量血液经过, 肾脏易受循环中的游离脂肪酸 (free fatty acid, FFA) 影响^[30-31]。血液中的FFA主要来源于食物中脂质的吸收或白色脂肪组织中甘油三酯 (triglyceride, TG) 的分解。脂肪组织中的甘油三酯脂肪酶 (adipose triglyceride lipase, ATGL) 催化脂肪细胞中TG水解的初始步骤, ATGL缺失小鼠的肾脏TG水平显著升高, 并伴有蛋白尿^[32]。细胞对FFA的摄取能力在很大程度上取决于血浆FA浓度以及膜结合脂肪酸转运蛋白 (fatty acid transporter proteins, FATPs) 和分化抗原36 (cluster of differentiation, CD36) 的表达水平^[33]。FATP2的表达减少会引起TECs脂质含量降低, 并抑制细胞凋亡^[34-35]。在肾脏疾病的发生发展中, CD36表达增加会导致脂质过载从而产生脂毒性^[36]。体内研究发现, CD36缺失的小鼠能够抵抗高脂饮食 (high-fat diet, HFD) 引起的肾脏脂质蓄积和炎性细胞浸润^[37]。

基于肾脏细胞能量代谢的研究表明, 肾损伤过程伴随着内皮细胞、足细胞和近端小管上皮细胞的脂肪生成增加^[38-39]。胆固醇调节元件结合蛋白1 (sterol-regulatory element binding proteins, SREBP-1) 和脂肪酸合成酶 (fatty acid synthase, FAS) 是FA从头合成的关键调节因子和限速酶。糖尿病肾病 (diabetic nephropathy, DN) 大鼠的肾组织以及棕

桐酸 (palmitic acid, PA) 诱导的 HK-2 细胞中 SREBP-1 和 FAS 的表达均显著升高; 抑制 SREBP-1、FAS 表达可以减轻 DN 大鼠肾小管异位脂质沉积, 并改善 PA 诱导的 TECs 脂质积累^[40]。另一关键的脂质合成调节因子糖应答元件结合蛋白 (carbohydrate response element binding protein, ChREBP) 的过表达会驱动 HK-2 细胞中活性氧 (reactive oxygen species, ROS) 产生^[41]。

脂质分解减少也是肾脏脂代谢失衡的关键因素。ATGL 和激素敏感脂肪酶 (hormone-sensitive lipase, HSL) 是 TG 分解的关键酶, 上调肾脏中 ATGL 和 HSL 的表达可以减轻肾小管中的脂质累积^[40]。脂肪酸氧化 (fatty acid oxidation, FAO) 受阻也会导致 TECs 内脂质积累, 并能促进上皮-间质转化 (epithelial-mesenchymal transition, EMT) 和炎症^[42]。FFA 的 β 氧化是近端肾小管细胞 ATP 产物的主要来源, 抑制脂肪酸 β 氧化关键酶肉毒碱棕榈酰转移酶 1 α (carnitine palmitoyltransferase 1 α , CPT1 α), 造成近端肾小管细胞内脂质沉积增加、ATP 耗竭和细胞死亡。过氧化物酶体增殖物激活 PPAR 活受体 α (peroxisome proliferators-activated receptors, PPAR α) 的激活可诱导线粒体 FAO 相关基因的转录, 如 CPT1 α 和酰基辅酶 a 氧化酶 1 (acyl-coenzyme A oxidase 1, ACOX1), 从而减少脂质蓄积^[43]。

综上所述, 肾脏脂质沉积是 FA 摄取、脂质合成和分解之间不平衡的结果, 想要改善肾脏的脂质累积则需要打破这种不平衡状态, 重新建立 3 条脂代谢途径之间的稳态, 为肾脏正常功能的维持提供能量。脂质稳态的破坏会促进组织和器官的病理变化, 从而导致生物衰老和与年龄相关的疾病。

3 脂代谢紊乱引发肾脏衰老的机制

3.1 脂肪酸摄取、脂质合成与肾脏衰老

FA 摄取和脂质合成异常增多是肾脏衰老的风险因素。D-半乳糖构建的衰老动物模型的肾组织中 CD36 的表达远高于正常组, CD36 的缺失则能够减轻衰老相关的肾损伤^[44]。调节 FA 从头合成的关键转录因子 SREBP 的激活不仅会造成肾损伤, 还会引发肾脏衰老^[45]。长期 HFD 造成机体血脂异常的同时也可加快肾脏细胞的衰老^[46]。研究表明, HFD 促进肾脏 p16、p19 和 p53 的表达以及 SA- β -Gal 的活性, 诱导肾脏衰老^[22]。茶多酚中最有效的活性成分表没食子儿茶素没食子酸酯给药导致肥胖大

鼠模型的血脂水平降低, 血清中白介素-6 (interleukin-6, IL-6)、肿瘤坏死因子- α (tumor necrosis factor- α , TNF- α) 等 SASP 因子减少, 并能够改善肾损伤以及延长动物模型的使用寿命^[47]。

3.2 脂质分解与肾脏衰老

肾脏功能的维持主要依赖于 FAO 途径产生的能量^[48], 大多数的肾脏疾病模型中都能检测到 FAO 障碍, 其中就包括衰老的肾脏^[29]。研究发现, 衰老状态的 TECs 的 FAO 途径受到损害, 进而加重肾脏脂质积累及肾小管损伤^[49]。PPAR 活性的降低与肾脏疾病中的脂质代谢紊乱直接相关^[11], 衰老肾脏中 PPAR α 的转录活性和蛋白质水平降低, PPAR α / β 受损与衰老过程中的慢性炎症有关^[49]。PPAR α / β 的激活剂 MHY2013 可以显著增加 FAO 相关基因的表达, 改善衰老 SD 大鼠 TECs 的脂质紊乱, 并下调 TGF- β 等基因的表达^[50]。上述研究表明, FAO 障碍是肾脏衰老过程中脂质代谢失调的关键因素。而在非 FAO 障碍造成的脂质累积的动物模型中, 脂质过氧化则可能是造成肾脏细胞损伤和衰老的驱动因素。例如, 4-羟基壬烯醛 (4-hydroxynonenal, 4-HNE) 的染色结果显示, 18 月龄的小鼠肾脏中脂质过氧化水平明显高于 12 月龄的小鼠^[51]。

3.3 应激性损伤与肾脏衰老

脂质的持续蓄积产生脂毒性, 诱发氧化应激、内质网应激和线粒体功能障碍, 造成 TECs 损伤^[52-53]。大量研究表明, TECs 损伤和衰老会导致肾功能障碍, 与肾脏疾病的发病密切相关, 包括 DN 和肾脏衰老等^[10, 54-55]。抑制小鼠 TECs 内质网应激可减轻 DN 进程中细胞的过早衰老^[56]。氧化应激可引起 DNA 氧化损伤和线粒体功能障碍, 对 TECs 的衰老有很大的影响^[57]。研究指出, 醛固酮可以诱导 TECs 的氧化应激和衰老, 提高细胞的自噬则可通过改善氧化应激减轻细胞衰老^[58]。氧化应激可诱导脂质过氧化, 形成高度反应性和亲电性的化合物, 这些化合物攻击蛋白质中的游离氨基使其共价修饰, 并导致高级脂氧化终产物 (advanced lipoxidation endproducts, ALEs) 的产生^[59], 氧化应激诱导的 ALEs 形成与衰老有关^[60]。作为诱导 HK-2 细胞衰老的常用试剂, H₂O₂ 增加了氧化应激和导致线粒体功能障碍^[61]。线粒体是 ROS 产生的主要场所, 线粒体 ROS 可以激活 Jun-N 端激酶, 促进染色质片段向胞质释放以及 SASP 的分泌^[62]。同时, 小鼠线粒体中具有抗氧化作用的超氧化物歧化

酶 (superoxide dismutase 2, SOD2) 的缺失则会导致细胞衰老 [63]。

4 脂代谢紊乱与肾纤维化的发生发展密切相关

在肥胖相关的肾脏疾病和肾纤维化进展中可以观察到, 脂质过载会产生脂毒性, 同时产生 ROS、释放促炎和促纤维化因子以及导致细胞周期阻滞, 促进细胞凋亡 [64]。血浆中脂蛋白的含量对肾纤维化有很大的影响。CKD 患者往往伴随着高甘油三酯血症、高的低密度脂蛋白胆固醇 (low density lipoprotein cholesterol, LDL-C) 和低的高密度脂蛋白胆固醇 (high-density lipoprotein cholesterol, HDL-C) 水平 [65-66]。接受肾脏移植的 CKD 患者的 LDL 水平与移植肾的脂质含量和纤维化密切相关 [67]。氧化 LDL (ox-LDL) 促进 TECs 的 EMT 过程, 引起 ECM 积累, 最终导致肾间质纤维化的发生 [68]。同样地, LDL 受体、乙酰化 LDL 和 CD36 水平的升高与 DN 的进展和 eGFR 的降低有关 [69]。临床数据表明, HDL-C 水平升高能明显改善肾功能 [70-71]。补充 HDL 可以降低黏附分子表达, 减少炎性细胞浸润以及细胞氧化应激, 并减轻肾小管损伤和肾纤维化, 提高肾缺血再灌注动物模型的

eGFR [72]。对高血脂有显著疗效的他汀类药物可明显减轻受试者的肾损伤 [73]。

脂质代谢紊乱引起的脂毒性通过各种致病机制损害 TECs、足细胞和肾小管间质细胞, 导致肾功能不全 [74]。在小鼠纤维化模型中, miR-21 介导的 PPAR α 下调会导致肾纤维化和肾小管上皮损伤 [75]。过表达 CPT1 α 可以增强 TECs 的 FAO, 降低炎症因子的表达, 并显著减轻纤维化 [76]。同样有研究指出, SREBP-1/2/AMPK 途径可启动肾脏中脂肪生成, 并通过上调 CD36 的表达加快脂质摄取, 双重作用下肾脏中脂质发生堆积, 造成肾损伤和肾纤维化 [67]。芝麻油中的有效活性成分芝麻素给药可介导脂质代谢, 改善高脂血症大鼠模型中脂质代谢紊乱引起的肾损伤以及下调大鼠肾脏中 α -SMA 和四型胶原蛋白 (Col-IV) 的表达 [77]。

因此, 肾脏中脂质过量累积会导致脂代谢紊乱, 产生脂毒性。同时产生内质网应激, 细胞内的活性氧和线粒体活性氧增加。脂毒性会诱发 TECs 应激性损伤和衰老, 衰老 TECs 分泌的 SASP 因子 (包括促纤维化因子和促炎因子) 进一步加速其衰老以及炎症和 EMT 的发生, 促进成纤维细胞活化和 ECM 的分泌, 最终导致肾间质纤维化的发生 (图 1)。

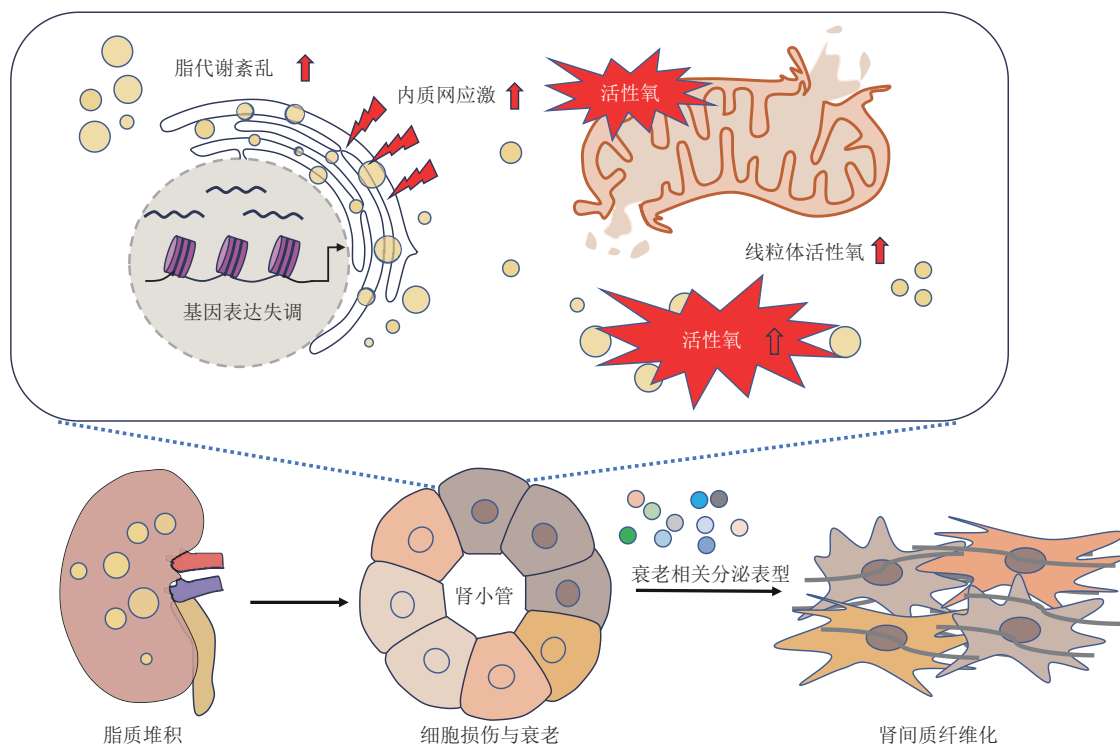


Fig. 1 Association of disorders of lipid metabolism with renal ageing and renal interstitial fibrosis

图1 脂代谢异常与肾脏衰老和肾间质纤维化的关系

5 基于脂代谢调控预防肾脏衰老与肾纤维化的潜在靶点和策略

5.1 潜在分子靶点类型

目前关于抑制甚至扭转肾间质纤维化的研究, 多数停留在实验动物阶段, 仅个别被应用到临床^[78-79]。延缓肾脏衰老的有效药物尚未见报道。现已明确脂质紊乱可以促进肾脏衰老与肾间质纤维化的发生发展。因此, 靶向脂质代谢相关酶、转运蛋白、转录因子有望成为预防或治疗肾间质纤维化的有效途径^[80]。

5.1.1 PPARs

PPARs 是配体激活的核受体/转录因子的一个亚家族, 属于核受体超家族^[81]。PPAR α 或 PPAR β 是细胞内脂质代谢的重要调节因子, 主要调控过氧化物酶体和线粒体 FAO 途径、FA 摄取和 TG 分解代谢相关基因的转录^[82]。老年大鼠肾脏中 PPAR α 和 FAO 相关酶的水平降低, 伴随着脂质堆积和肾纤维化的显著增加^[50]。研究表明, PPAR α 缺失造成的脂代谢失衡在衰老相关的肾纤维化发展中发挥重要作用。PPAR α 敲除的老年小鼠的肾脏表现出脂质积聚、损伤加重以及纤维化水平升高, miR-21 也可通过下调 PPAR α 表达加快小鼠模型的肾小管上皮损伤和纤维化^[49]。PPAR α/β 激活剂则能够减少细胞内脂质积聚并下调细胞中衰老和纤维化相关基因的表达, 如 *p65*、*IL-1*、*IL-6* 和 *TNF- α* 等^[50]。多酚类中药单体白藜芦醇通过调节 PPAR α 途径预防 HFD 小鼠肾脂毒性^[83]。研究表明, 核受体 PPAR γ 的活性随着年龄的增长而降低^[84]。可保护人体细胞免受氧化应激和衰老的影响^[85-87]。

5.1.2 PGC1 α

PGC1 α 是转录共激活因子家族的一员, 是 PPARs 等转录调控蛋白的共激活因子。它在能量代谢组织中广泛表达^[30], 参与调节能量代谢、线粒体生物发生以及其他生物过程^[88]。在 CKD 患者肾脏中, PGC1 α 表达下调, 其表达与 eGFR 呈正相关, 与纤维化呈负相关^[89]。肾小管特异性过表达 PGC1 α 可恢复线粒体含量并减轻纤维化^[48]。在衰老或发生纤维化肾脏中, PGC1 α 和相关转录因子的缺乏会导致 FAO 障碍、脂质堆积、过量 ROS 的产生、线粒体膜电位的丧失和线粒体损伤^[90]。研究表明, 沉默信息调节因子 1 (silent information regulator 1, Sirt1) 可以通过激活 PGC1 α 来改善线粒体的生物发生从而延长寿命^[91]。PGC1 α 还是

Notch 的靶基因, 上调 *PGC1 α* 基因表达可以预防 Notch 介导的肾损伤和纤维化^[89]。考虑到调控脂代谢和线粒体生物发生的重要作用, PGC1 α 是极具潜力的一个治疗肾脏疾病分子靶点。

5.1.3 CPT1/2

CPT 家族成员 CPT1 和 CPT2 位于线粒体外膜和内膜中, CPT1/2 是 FAO 的一种关键限速酶, 对脂质分解代谢至关重要^[92]。FA 进入线粒体是由 CPT1 介导的, 该酶的作用是使 FA 与肉碱结合。CPT1 的下调会抑制 FAO, 造成脂质堆积, 进而导致 CKD 和肾纤维化^[48]。在衰老相关的肾纤维化患者中, CPT1 α 的表达与纤维化程度呈负相关^[93]。研究表明, 敲除 CPT1 会降低近端肾小管上皮细胞的 FAO 和 ATP 产生能力, 加剧肾小管损伤和肾纤维化, 并减弱肾脏的修复能力^[42]。CPT1 的过表达则可以保护线粒体呼吸并防止叶酸诱导的肾病小鼠模型的肾小管细胞损伤^[94]。同样, 在单侧输尿管结扎或腺嘌呤诱导的肾纤维化小鼠模型的 TECs 中过表达 CPT1, 引起上皮细胞损伤减轻、炎症反应减弱以及纤维化标志物的表达减少^[76]。间充质干细胞细胞外小泡通过恢复 CPT1 α 的表达, 增强肾脏线粒体 FAO, 修复了线粒体的结构, 在减轻肾损伤和肾纤维化方面表现出优越的治疗效果^[75]。DN 小鼠模型中 CPT2 及其调控分子的表达较低, 细胞内脂质沉积较多^[95]。CPT2 的过表达使 FAO 和纤维化相关基因恢复到正常水平^[95], 表明靶向 CPT2 有助于预防和治疗肾纤维化。

5.1.4 TGF- β

TGF- β 超家族在衰老、组织稳态、纤维化疾病和免疫功能障碍等一系列病理变化中起重要的作用^[96]。参与 EMT 和 ECM 积累的众多通路, TGF- β 1 被认为在肾纤维化中发挥主要作用^[97]。TGF- β 1 已被确定为细胞和器官衰老的关键调节因子, TGF- β 信号传导以多种方式与衰老过程相互作用。TGF- β 信号传导的下游靶点包括许多参与衰老过程的调节因子, 如细胞增殖、细胞周期调节、ROS 产生、DNA 损伤修复、端粒调节和自噬^[98]。人们普遍认为脂毒性代谢物的生成通常与脂质积累同时发生, 这在肾脏衰老和肾间质纤维化的发病机制中起着关键作用。脂毒性易使肾脏产生过多的 ROS 和氧化应激, 可引起细胞坏死和凋亡^[99]。在 TGF- β 刺激的 TECs 中, 谷胱甘肽浓度降低, 脂质过氧化增强, 暗示了 TGF- β 也可能通过调节肾脏细胞的脂代谢过程促进肾纤维化进程^[100]。

5.1.5 Sirt1

肾脏中 Sirt1 水平降低与肾功能减退有关^[87]。在肥胖的 DN 大鼠模型中, 饮食限制 (dietary restriction, DR) 会上调 Sirt1 的表达, 加强抗炎作用, 改善自噬失调, 从而改善肾损伤^[101]。Sirt1 的过表达还能降低 ROS、FN 和 TGF- β 1 水平^[102]。黄芩苷可以通过 Sirt1/AMPK/HNF4A 途径缓解 *db/db* 小鼠肾脏的脂代谢异常, 发挥肾脏保护作用^[103]。此外, Sirt1 对氧化应激介导的肾纤维也有重要的作用^[102], 同样的结果在 HK-2 细胞中也得到证实^[104]。Sirt1 是一种抗衰老分子^[105]。衰老过程中 Sirt1 的表达水平与线粒体生物发生保持一致^[106]。骨髓来源的间充质干细胞通过 Sirt1/Parkin 轴促进 TECs 的自噬, 改善脓毒症诱导的急性肾损伤^[107]。Sirt1 过表达和 SRT1720 治疗均降低了肾脏脂质含量和脂肪生成、氧化应激和炎症标志物的表达, 并在一定程度上减轻肾间质纤维化^[108]。白藜芦醇给药能通过激活 Sirt1 防止 HFD 小鼠高糖高脂引发的肾脏损伤和细胞衰老^[109]。

5.2 相关治疗策略

5.2.1 运动干预和热量限制

运动可以降低 BMI、腰围和脂肪量^[110]。定期运动可以降低 ROS 和血清糖基化终产物 (advanced glycation end product, AGE) 水平, 减轻衰老引起的氧化应激以及 p16、p21 的表达, 从而达到延缓衰老, 延长寿命的目的^[111]。AGE 是指在非酶促条件下, 蛋白质、氨基酸、脂类或核酸等大分子物质的游离氨基与还原糖的醛基经过缩合、重排、裂解、氧化修饰后产生的一组稳定的终末产物, 其过量产生会引发的细胞功能改变和机体病变, 坚持运动则能够减少这一情况的发生。在 HFD 小鼠模型中, 运动减少了衰老细胞的比率以及 SASP 因子的分泌, 并改善了机体功能^[112]。运动可以增强端粒酶逆转录酶基因的表达和端粒酶活性, 减轻端粒的磨损, 从而延缓细胞衰老^[113]。

鉴于 HFD 可以加速肾脏衰老, 研究人员进一步探究了减少脂质或者热量的摄入对衰老的作用, 发现 DR, 即是在不挨饿的情况下减少能量的摄入, 可以延迟健康寿命, 并且降低衰老相关疾病的发病率^[114]。长期以来, DR 被认为可以延长从苍蝇到灵长类动物等物种的寿命, 对细胞衰老机制的广泛影响。与短期 DR 相比, 长期 DR 对于延缓肾脏衰老具有更佳的效果^[115]。DR 对 CKD 具有保护作用, 可恢复肾脏功能, 降低 CKD 发病率, 提高生存

率^[116]。短期 DR 能够通过增加自噬活性和降低氧化应激来对肾脏衰老起到保护作用; 长期 DR 可以改善衰老肾脏的线粒体受损、氧化应激和纤维化^[117]。老年 DR 组的肾脏中 NF- κ B 的磷酸化水平显著下降以及 SASP 尤其是细胞因子、趋化因子和生长因子表达减少^[24], 提示 DR 通过减少降低 NF- κ B 的转录活性来抑制 SASP 因子分泌, 从而延缓肾脏衰老。

5.2.2 针对SASP因素的新兴干预措施

SASP 因子及其相关受体的分泌可引发肾脏衰老和肾间质纤维化^[118]。SASP 治疗主要包括以下 3 种途径: a. 通过调控转录因子表达或活性阻断 SASP 的前信号通路; b. 使用蛋白酶抑制 SASP 的生物活性; c. 抑制特定的 SASP 因子或其受体。例如, 用 shRNA 降低 NF- κ B 的表达已被证明可以阻止衰老细胞中 IL-6、IL-8、CXCL1 和 ICAM-1 的分泌^[119]。p38 抑制剂可以通过抑制 IL-6 表达来减轻肾纤维化^[120]。胡桃素给药通过调节 NF- κ B 信号传导改善 HFD 小鼠肾脏中的脂质沉积和炎症^[121]。柑橘皮中的有效活性成分川皮苷给药可以通过抑制 IL-6 和 TGF- β 来减轻肾纤维化^[122]。TGF- β 在纤维化过程中起着重要作用, 针对 TGF- β 途径的各种方法已经出现, 如特异性抗体、可溶性受体、TGF- β 结合蛋白、TGF- β 受体的小分子抑制剂, 以及相关的 microRNAs^[123]。然而, 仅阻断 TGF- β 1 信号在 CKD 中的抗纤维化效果并不理想^[124], 提示可能多个 SASP 因子在肾间质纤维化和肾脏衰老过程中共同起作用。

综上所述, 脂质过度累积会细胞脂毒性和 ROS 含量增加, 引发氧化应激、线粒体功能障碍、内质网应激等, 致使肾脏细胞衰老和肾间质纤维化发生。调节 FA 摄取、脂质合成与分解的平衡, 改善脂代谢紊乱, 有助于延缓肾脏衰老和减轻肾间质纤维化。鉴于肾间质纤维化和肾脏衰老带来的后果非常严重, 扭转肾间质纤维化和延缓肾脏衰老药物的成功研发和应用于临床成为极其期待的事情。但目前对肾纤维化治疗靶点的研究较为缺乏, 仍需要对治疗肾纤维化的靶点进行临床试验, 以评估其在患者中的安全性和有效性。此外, 许多炎症和纤维化因子也是 SASP 成分, 其表达不足以识别细胞衰老。鉴于现有的衰老指标可能因疾病和器官而异^[125], 因此需要更高灵敏度和特异性的衰老检测方法, 脂代谢途径的关键酶和调控蛋白有望成为改善肾脏衰老和肾间质纤维化的潜在靶点。从脂代谢

的角度, 深入挖掘肾脏衰老以及肾间质纤维化的分子调控机制, 寻找更多更好的治疗靶点, 筛选出更多的小分子药物, 对于延缓肾脏衰老以及治疗肾间质纤维化具有重要的意义以及临床使用价值。

参 考 文 献

- [1] Grimley Evans J. Ageing and medicine. *J Intern Med*, 2000, **247**(2): 159-167
- [2] Soegiarto G, Purnomosari D. Challenges in the vaccination of the elderly and strategies for improvement. *Pathophysiology*, 2023, **30**(2): 155-173
- [3] Fang E F, Scheibye-Knudsen M, Jahn H J, *et al.* A research agenda for aging in China in the 21st century. *Ageing Res Rev*, 2015, **24**(PtB): 197-205
- [4] Huang R, Fu P, Ma L. Kidney fibrosis: from mechanisms to therapeutic medicines. *Signal Transduct Target Ther*, 2023, **8**(1): 129
- [5] Widjaja A A, Viswanathan S, Shekeran S G, *et al.* Targeting endogenous kidney regeneration using anti-IL11 therapy in acute and chronic models of kidney disease. *Nat Commun*, 2022, **13**(1): 7497
- [6] Nitta K, Okada K, Yanai M, *et al.* Aging and chronic kidney disease. *Kidney Blood Press Res*, 2013, **38**(1): 109-120
- [7] Bridges C C, Zalups R K. The aging kidney and the nephrotoxic effects of mercury. *J Toxicol Environ Health B Crit Rev*, 2017, **20**(2): 55-80
- [8] Fang Y, Gong A Y, Haller S T, *et al.* The ageing kidney: molecular mechanisms and clinical implications. *Ageing Res Rev*, 2020, **63**: 101151
- [9] Santin Y, Lluel P, Rischmann P, *et al.* Cellular senescence in renal and urinary tract disorders. *Cells*, 2020, **9**(11): 2420
- [10] Gekle M. Kidney and aging - a narrative review. *Exp Gerontol*, 2017, **87**(PtB): 153-155
- [11] Speeckaert M M, Vanfraechem C, Speeckaert R, *et al.* Peroxisome proliferator-activated receptor agonists in a battle against the aging kidney. *Ageing Res Rev*, 2014, **14**: 1-18
- [12] Sis B, Tasanarong A, Khoshjou F, *et al.* Accelerated expression of senescence associated cell cycle inhibitor p16INK4A in kidneys with glomerular disease. *Kidney Int*, 2007, **71**(3): 218-226
- [13] Docherty M H, O'sullivan E D, Bonventre J V, *et al.* Cellular senescence in the kidney. *J Am Soc Nephrol*, 2019, **30**(5): 726-736
- [14] Baker D J, Childs B G, Durik M, *et al.* Naturally occurring p16 (Ink4a) -positive cells shorten healthy lifespan. *Nature*, 2016, **530**(7589): 184-189
- [15] Humphreys B D. Mechanisms of renal fibrosis. *Annu Rev Physiol*, 2018, **80**: 309-326
- [16] Djurdjaj S, Boor P. Cellular and molecular mechanisms of kidney fibrosis. *Mol Aspects Med*, 2019, **65**: 16-36
- [17] Peek J L, Wilson M H. Cell and gene therapy for kidney disease. *Nat Rev Nephrol*, 2023, **19**(7): 451-462
- [18] Lovisa S, Lebleu V S, Tampe B, *et al.* Epithelial-to-mesenchymal transition induces cell cycle arrest and parenchymal damage in renal fibrosis. *Nat Med*, 2015, **21**(9): 998-1009
- [19] Yang C, Chen X C, Li Z H, *et al.* SMAD3 promotes autophagy dysregulation by triggering lysosome depletion in tubular epithelial cells in diabetic nephropathy. *Autophagy*, 2021, **17**(9): 2325-2344
- [20] Rayego-Mateos S, Valdivielso J M. New therapeutic targets in chronic kidney disease progression and renal fibrosis. *Expert Opin Ther Targets*, 2020, **24**(7): 655-670
- [21] Ferenbach D A, Bonventre J V. Mechanisms of maladaptive repair after AKI leading to accelerated kidney ageing and CKD. *Nat Rev Nephrol*, 2015, **11**(5): 264-276
- [22] Kim S R, Jiang K, Ogrodnik M, *et al.* Increased renal cellular senescence in murine high-fat diet: effect of the senolytic drug quercetin. *Transl Res*, 2019, **213**: 112-123
- [23] Dong D, Cai G Y, Ning Y C, *et al.* Alleviation of senescence and epithelial-mesenchymal transition in aging kidney by short-term caloric restriction and caloric restriction mimetics via modulation of AMPK/mTOR signaling. *Oncotarget*, 2017, **8**(10): 16109-16121
- [24] Wang W, Cai G, Chen X. Dietary restriction delays the secretion of senescence associated secretory phenotype by reducing DNA damage response in the process of renal aging. *Exp Gerontol*, 2018, **107**: 4-10
- [25] Clements M E, Chaber C J, Ledbetter S R, *et al.* Increased cellular senescence and vascular rarefaction exacerbate the progression of kidney fibrosis in aged mice following transient ischemic injury. *PLoS One*, 2013, **8**(8): e70464
- [26] Yang L, Besschetnova T Y, Brooks C R, *et al.* Epithelial cell cycle arrest in G2/M mediates kidney fibrosis after injury. *Nat Med*, 2010, **16**(5): 535-543, 1p following 143
- [27] Luo C, Zhou S, Zhou Z, *et al.* Wnt9a promotes renal fibrosis by accelerating cellular senescence in tubular epithelial cells. *J Am Soc Nephrol*, 2018, **29**(4): 1238-1256
- [28] Houten S M, Violante S, Ventura F V, *et al.* The biochemistry and physiology of mitochondrial fatty acid beta-oxidation and its genetic disorders. *Annu Rev Physiol*, 2016, **78**: 23-44
- [29] Chung K W. Advances in understanding of the role of lipid metabolism in aging. *Cells*, 2021, **10**(4): 880
- [30] Lin P H, Duann P. Dyslipidemia in kidney disorders: perspectives on mitochondria homeostasis and therapeutic opportunities. *Front Physiol*, 2020, **11**: 1050
- [31] Castro B B A, Foresto-Neto O, Saraiva-Camara N O, *et al.* Renal lipotoxicity: insights from experimental models. *Clin Exp Pharmacol Physiol*, 2021, **48**(12): 1579-1588
- [32] Chen W, Jiang Y, Han J, *et al.* Atgl deficiency induces podocyte apoptosis and leads to glomerular filtration barrier damage. *FEBS J*, 2017, **284**(7): 1070-1081
- [33] Rada P, Gonzalez-Rodriguez A, Garcia-Monzon C, *et al.* Understanding lipotoxicity in NAFLD pathogenesis: is CD36 a key driver?. *Cell Death Dis*, 2020, **11**(9): 802
- [34] Khan S, Cabral P D, Schilling W P, *et al.* Kidney proximal tubule

- lipoapoptosis is regulated by fatty acid transporter-2 (FATP2). *J Am Soc Nephrol*, 2018, **29**(1): 81-91
- [35] Chen Y, Yan Q, Lv M, *et al.* Involvement of FATP2-mediated tubular lipid metabolic reprogramming in renal fibrogenesis. *Cell Death Dis*, 2020, **11**(11): 994
- [36] Li X, Zhang T, Geng J, *et al.* Advanced oxidation protein products promote lipotoxicity and tubulointerstitial fibrosis via CD36/beta-catenin pathway in diabetic nephropathy. *Antioxid Redox Signal*, 2019, **31**(7): 521-538
- [37] Yang X, Okamura D M, Lu X, *et al.* CD36 in chronic kidney disease: novel insights and therapeutic opportunities. *Nat Rev Nephrol*, 2017, **13**(12): 769-781
- [38] Gai Z, Wang T, Visentin M, *et al.* Lipid accumulation and chronic kidney disease. *Nutrients*, 2019, **11**(4): 7-22
- [39] Opazo-Rios L, Mas S, Marin-Royo G, *et al.* Lipotoxicity and diabetic nephropathy: novel mechanistic insights and therapeutic opportunities. *Int J Mol Sci*, 2020, **21**(7): 2632
- [40] Su K, Yi B, Yao B Q, *et al.* Liraglutide attenuates renal tubular ectopic lipid deposition in rats with diabetic nephropathy by inhibiting lipid synthesis and promoting lipolysis. *Pharmacol Res*, 2020, **156**: 104778
- [41] Suzuki S, Yokoyama A, Noro E, *et al.* Expression and pathophysiological significance of carbohydrate response element binding protein (ChREBP) in the renal tubules of diabetic kidney. *Endocr J*, 2020, **67**(3): 335-345
- [42] Calle P, Torrico S, Munoz A, *et al.* CPT1a downregulation protects against cholesterol-induced fibrosis in tubular epithelial cells by downregulating TGFbeta-1 and inflammasome. *Biochem Biophys Res Commun*, 2019, **517**(4): 715-721
- [43] Kersten S, Stienstra R. The role and regulation of the peroxisome proliferator activated receptor alpha in human liver. *Biochimie*, 2017, **136**: 75-84
- [44] Peng J, Ren X F, Yang C, *et al.* Effects of inflammatory response on renal function and TGF-beta1 pathway of rats with aging-related kidney damage by upregulating the expression of CD36. *Eur Rev Med Pharmacol Sci*, 2020, **24**(17): 8957-8967
- [45] Jiang T, Liebman S E, Lucia M S, *et al.* Role of altered renal lipid metabolism and the sterol regulatory element binding proteins in the pathogenesis of age-related renal disease. *Kidney Int*, 2005, **68**(6): 2608-2620
- [46] Seidell J C. Dietary fat and obesity: an epidemiologic perspective. *Am J Clin Nutr*, 1998, **67**(3 Suppl): 546S-550S
- [47] Yuan H, Li Y, Ling F, *et al.* The phytochemical epigallocatechin gallate prolongs the lifespan by improving lipid metabolism, reducing inflammation and oxidative stress in high-fat diet-fed obese rats. *Aging Cell*, 2020, **19**(9): e13199
- [48] Kang H M, Ahn S H, Choi P, *et al.* Defective fatty acid oxidation in renal tubular epithelial cells has a key role in kidney fibrosis development. *Nat Med*, 2015, **21**(1): 37-46
- [49] Chung K W, Lee E K, Lee M K, *et al.* Impairment of PPARalpha and the fatty acid oxidation pathway aggravates renal fibrosis during aging. *J Am Soc Nephrol*, 2018, **29**(4): 1223-1237
- [50] Chung K W, Ha S, Kim S M, *et al.* PPARalpha/beta activation alleviates age-associated renal fibrosis in sprague dawley rats. *J Gerontol A Biol Sci Med Sci*, 2020, **75**(3): 452-458
- [51] Marquez-Exposito L, Tejedor-Santamaria L, Valentijn F A, *et al.* Oxidative stress and cellular senescence are involved in the aging kidney. *Antioxidants (Basel)*, 2022, **11**(2): 301
- [52] Lhotak S, Sood S, Brimble E, *et al.* ER stress contributes to renal proximal tubule injury by increasing SREBP-2-mediated lipid accumulation and apoptotic cell death. *Am J Physiol Renal Physiol*, 2012, **303**(2): F266-278
- [53] Nishi H, Higashihara T, Inagi R. Lipotoxicity in kidney, heart, and skeletal muscle dysfunction. *Nutrients*, 2019, **11**(7): 1664
- [54] He S, Sharpless N E. Senescence in health and disease. *Cell*, 2017, **169**(6): 1000-1011
- [55] Xiong Y, Zhou L. The signaling of cellular senescence in diabetic nephropathy. *Oxid Med Cell Longev*, 2019, **2019**: 7495629
- [56] Liu J, Yang J R, Chen X M, *et al.* Impact of ER stress-regulated ATF4/p16 signaling on the premature senescence of renal tubular epithelial cells in diabetic nephropathy. *Am J Physiol Cell Physiol*, 2015, **308**(8): C621-C630
- [57] Schmitt R, Melk A. Molecular mechanisms of renal aging. *Kidney Int*, 2017, **92**(3): 569-579
- [58] Cao J Y, Ling L L, Ni W J, *et al.* Autophagosome protects proximal tubular cells from aldosterone-induced senescence through improving oxidative stress. *Ren Fail*, 2021, **43**(1): 556-565
- [59] Vistoli G, De Maddis D, Cipak A, *et al.* Advanced glycoxidation and lipoxidation end products (AGEs and ALEs): an overview of their mechanisms of formation. *Free Radic Res*, 2013, **47**(Suppl 1): 3-27
- [60] Ahmad S, Siddiqui Z, Rehman S, *et al.* A glycation angle to look into the diabetic vasculopathy: cause and cure. *Curr Vasc Pharmacol*, 2017, **15**(4): 352-364
- [61] Small D M, Bennett N C, Roy S, *et al.* Oxidative stress and cell senescence combine to cause maximal renal tubular epithelial cell dysfunction and loss in an *in vitro* model of kidney disease. *Nephron Exp Nephrol*, 2012, **122**(3-4): 123-130
- [62] Vizioli M G, Liu T, Miller K N, *et al.* Mitochondria-to-nucleus retrograde signaling drives formation of cytoplasmic chromatin and inflammation in senescence. *Genes Dev*, 2020, **34**(5-6): 428-445
- [63] Velarde M C, Flynn J M, Day N U, *et al.* Mitochondrial oxidative stress caused by Sod2 deficiency promotes cellular senescence and aging phenotypes in the skin. *Aging (Albany NY)*, 2012, **4**(1): 3-12
- [64] Izquierdo-Lahuerta A, Martinez-Garcia C, Medina-Gomez G. Lipotoxicity as a trigger factor of renal disease. *J Nephrol*, 2016, **29**(5): 603-610
- [65] Catapano A L, Graham I, De Backer G, *et al.* 2016 ESC/EAS guidelines for the management of dyslipidaemias. *Eur Heart J*, 2016, **37**(39): 2999-3058
- [66] Ferro C J, Mark P B, Kanbay M, *et al.* Lipid management in patients with chronic kidney disease. *Nat Rev Nephrol*, 2018, **14**(12): 727-749

- [67] Lin Y C, Wang J C, Wu M S, *et al.* Nifedipine exacerbates lipogenesis in the kidney *via* KIM-1, CD36, and SREBP upregulation: implications from an animal model for human study. *Int J Mol Sci*, 2020, **21**(12): 4359
- [68] Sung P H, Cheng B C, Hsu T W, *et al.* Oxidized-LDL deteriorated the renal residual function and parenchyma in CKD rat through upregulating epithelial mesenchymal transition and extracellular matrix-mediated tubulointerstitial fibrosis-pharmacomodulation of rosuvastatin. *Antioxidants (Basel)*, 2022, **11**(12): 2465
- [69] Chen S, Chen J, Li S, *et al.* High-fat diet-induced renal proximal tubular inflammatory injury: emerging risk factor of chronic kidney disease. *Front Physiol*, 2021, **12**: 786599
- [70] Lanktree M B, Theriault S, Walsh M, *et al.* HDL cholesterol, LDL cholesterol, and triglycerides as risk factors for CKD: a mendelian randomization study. *Am J Kidney Dis*, 2018, **71**(2): 166-172
- [71] Liu H M, Hu Q, Zhang Q, *et al.* Causal effects of genetically predicted cardiovascular risk factors on chronic kidney disease: a two-sample mendelian randomization study. *Front Genet*, 2019, **10**: 415
- [72] Moreira R S, Irigoyen M, Sanches T R, *et al.* Apolipoprotein A-I mimetic peptide 4F attenuates kidney injury, heart injury, and endothelial dysfunction in sepsis. *Am J Physiol Regul Integr Comp Physiol*, 2014, **307**(5): R514-R524
- [73] Steinmetz O M, Panzer U, Stahl R A, *et al.* Statin therapy in patients with chronic kidney disease: to use or not to use. *Eur J Clin Invest*, 2006, **36**(8): 519-527
- [74] Wanders R J, Waterham H R, Ferdinandusse S. Metabolic interplay between peroxisomes and other subcellular organelles including mitochondria and the endoplasmic reticulum. *Front Cell Dev Biol*, 2015, **3**: 83
- [75] Gao Z, Zhang C, Peng F, *et al.* Hypoxic mesenchymal stem cell-derived extracellular vesicles ameliorate renal fibrosis after ischemia-reperfusion injury by restoring CPT1A mediated fatty acid oxidation. *Stem Cell Res Ther*, 2022, **13**(1): 191
- [76] Miguel V, Tituana J, Herrero J I, *et al.* Renal tubule Cpt1a overexpression protects from kidney fibrosis by restoring mitochondrial homeostasis. *J Clin Invest*, 2021, **131**(5): e140695
- [77] Zhang R, Yu Y, Deng J, *et al.* Sesamin ameliorates high-fat diet-induced dyslipidemia and kidney injury by reducing oxidative stress. *Nutrients*, 2016, **8**(5): 276
- [78] Remuzzi G, Cattaneo D, Perico N. The aggravating mechanisms of aldosterone on kidney fibrosis. *J Am Soc Nephrol*, 2008, **19**(8): 1459-1462
- [79] Tu X, Chen X, Xie Y, *et al.* Anti-inflammatory renoprotective effect of clopidogrel and irbesartan in chronic renal injury. *J Am Soc Nephrol*, 2008, **19**(1): 77-83
- [80] Chen Y Y, Chen X G, Zhang S. Druggability of lipid metabolism modulation against renal fibrosis. *Acta Pharmacol Sin*, 2022, **43**(3): 505-519
- [81] Michalik L, Auwerx J, Berger J P, *et al.* International union of pharmacology. LXI. peroxisome proliferator-activated receptors. *Pharmacol Rev*, 2006, **58**(4): 726-741
- [82] Tong L, Wang L, Yao S, *et al.* PPARdelta attenuates hepatic steatosis through autophagy-mediated fatty acid oxidation. *Cell Death Dis*, 2019, **10**(3): 197
- [83] Zhou Y, Lin S, Zhang L, *et al.* Resveratrol prevents renal lipotoxicity in high-fat diet-treated mouse model through regulating PPAR-alpha pathway. *Mol Cell Biochem*, 2016, **411**(1-2): 143-150
- [84] Wang P, Li B, Cai G, *et al.* Activation of PPAR-gamma by pioglitazone attenuates oxidative stress in aging rat cerebral arteries through upregulating UCP2. *J Cardiovasc Pharmacol*, 2014, **64**(6): 497-506
- [85] Briganti S, Flori E, Bellei B, *et al.* Modulation of PPARgamma provides new insights in a stress induced premature senescence model. *PLoS One*, 2014, **9**(8): e104045
- [86] Chau B N, Xin C, Hartner J, *et al.* MicroRNA-21 promotes fibrosis of the kidney by silencing metabolic pathways. *Sci Transl Med*, 2012, **4**(121): 121ra118
- [87] Liu H J, Miao H, Yang J Z, *et al.* Deciphering the role of lipoproteins and lipid metabolic alterations in ageing and ageing-associated renal fibrosis. *Ageing Res Rev*, 2023, **85**: 101861
- [88] Fontecha-Barriuso M, Martin-Sanchez D, Martinez-Moreno J M, *et al.* The role of PGC-1alpha and mitochondrial biogenesis in kidney diseases. *Biomolecules*, 2020, **10**(2): 347
- [89] Han S H, Wu M Y, Nam B Y, *et al.* PGC-1alpha protects from Notch-induced kidney fibrosis development. *J Am Soc Nephrol*, 2017, **28**(11): 3312-3322
- [90] Lee G, Uddin M J, Kim Y, *et al.* PGC-1alpha, a potential therapeutic target against kidney aging. *Ageing Cell*, 2019, **18**(5): e12994
- [91] Yuan Y, Cruzat V F, Newsholme P, *et al.* Regulation of SIRT1 in aging: roles in mitochondrial function and biogenesis. *Mech Ageing Dev*, 2016, **155**: 10-21
- [92] Song X, Du Z, Yao Z, *et al.* Rhein improves renal fibrosis by restoring Cpt1a-mediated fatty acid oxidation through SirT1/STAT3/twist1 pathway. *Molecules*, 2022, **27**(7): 2344
- [93] Braun F, Rinschen M M, Bartels V, *et al.* Altered lipid metabolism in the aging kidney identified by three layered omic analysis. *Ageing (Albany NY)*, 2016, **8**(3): 441-457
- [94] Yuan Q, Lv Y, Ding H, *et al.* CPT1alpha maintains phenotype of tubules *via* mitochondrial respiration during kidney injury and repair. *Cell Death Dis*, 2021, **12**(8): 792
- [95] Warfel J D, Bermudez E M, Mendoza T M, *et al.* Mitochondrial fat oxidation is essential for lipid-induced inflammation in skeletal muscle in mice. *Sci Rep*, 2016, **6**: 37941
- [96] David C J, Massague J. Contextual determinants of TGFbeta action in development, immunity and cancer. *Nat Rev Mol Cell Biol*, 2018, **19**(7): 419-435
- [97] Wang Y Y, Jiang H, Pan J, *et al.* Macrophage-to-myofibroblast transition contributes to interstitial fibrosis in chronic renal allograft injury. *J Am Soc Nephrol*, 2017, **28**(7): 2053-2067
- [98] Bao C, Yang Z, Cai Q, *et al.* Incremental load training improves renal fibrosis by regulating the TGF-beta1/TAK1/MKK3/

- p38MAPK signaling pathway and inducing the activation of autophagy in aged mice. *Int J Mol Med*, 2019, **44**(5): 1677-1686
- [99] Mikula-Pietrasik J, Rutecki S, Ksiazek K. The functional multipotency of transforming growth factor beta signaling at the intersection of senescence and cancer. *Cell Mol Life Sci*, 2022, **79**(4): 196
- [100] Kim S, Kang S W, Joo J, *et al.* Characterization of ferroptosis in kidney tubular cell death under diabetic conditions. *Cell Death Dis*, 2021, **12**(2): 160
- [101] Kitada M, Takeda A, Nagai T, *et al.* Dietary restriction ameliorates diabetic nephropathy through anti-inflammatory effects and regulation of the autophagy *via* restoration of Sirt1 in diabetic Wistar fatty (fa/fa) rats: a model of type 2 diabetes. *Exp Diabetes Res*, 2011, **2011**: 908185
- [102] Huang K, Huang J, Xie X, *et al.* Sirt1 resists advanced glycation end products-induced expressions of fibronectin and TGF-beta1 by activating the Nrf2/ARE pathway in glomerular mesangial cells. *Free Radic Biol Med*, 2013, **65**: 528-540
- [103] Zhang Y, Yao H, Li C, *et al.* Gandi Capsule improved podocyte lipid metabolism of diabetic nephropathy mice through SIRT1/AMPK/HNF4A Pathway. *Oxid Med Cell Longev*, 2022, **2022**: 6275505
- [104] Xue M, Li Y, Hu F, *et al.* High glucose up-regulates microRNA-34a-5p to aggravate fibrosis by targeting SIRT1 in HK-2 cells. *Biochem Biophys Res Commun*, 2018, **498**(1): 38-44
- [105] Waldman M, Cohen K, Yadin D, *et al.* Regulation of diabetic cardiomyopathy by caloric restriction is mediated by intracellular signaling pathways involving 'SIRT1 and PGC-1alpha'. *Cardiovasc Diabetol*, 2018, **17**(1): 111
- [106] Chen G, Li X. The decreased SIRT1 level may account for the lipid profile in chronic kidney disease. *J Biol Res (Thessalon)*, 2019, **26**: 9
- [107] Guo J, Wang R, Liu D. Bone marrow-derived mesenchymal stem cells ameliorate sepsis-induced acute kidney injury by promoting mitophagy of renal tubular epithelial cells *via* the SIRT1/Parkin axis. *Front Endocrinol (Lausanne)*, 2021, **12**: 639165
- [108] Nguyen L T, Mak C H, Chen H, *et al.* SIRT1 attenuates kidney disorders in male offspring due to maternal high-fat diet. *Nutrients*, 2019, **11**(1): 146
- [109] Zhang N, Li Z, Xu K, *et al.* Resveratrol protects against high-fat diet induced renal pathological damage and cell senescence by activating SIRT1. *Biol Pharm Bull*, 2016, **39**(9): 1448-1454
- [110] Englund D A, Sakamoto A E, Fritsche C M, *et al.* Exercise reduces circulating biomarkers of cellular senescence in humans. *Aging Cell*, 2021, **20**(7): e13415
- [111] Lee J Y, Paik I Y, Kim J Y. Voluntary exercise reverses immune aging induced by oxidative stress in aging mice. *Exp Gerontol*, 2019, **115**: 148-154
- [112] Schafer M J, White T A, Evans G, *et al.* Exercise prevents diet-induced cellular senescence in adipose tissue. *Diabetes*, 2016, **65**(6): 1606-1615
- [113] Denham J, Sellami M. Exercise training increases telomerase reverse transcriptase gene expression and telomerase activity: a systematic review and meta-analysis. *Ageing Res Rev*, 2021, **70**: 101411
- [114] Fontana L, Partridge L, Longo V D. Extending healthy life span—from yeast to humans. *Science*, 2010, **328**(5976): 321-326
- [115] Wang W J, Cai G Y, Ning Y C, *et al.* Hydrogen sulfide mediates the protection of dietary restriction against renal senescence in aged F344 rats. *Sci Rep*, 2016, **6**: 30292
- [116] Singh G, Krishan P. Dietary restriction regimens for fighting kidney disease: insights from rodent studies. *Exp Gerontol*, 2019, **128**: 110738
- [117] Chiang C H, Li S J, Zhang T R, *et al.* Long-term dietary restriction ameliorates ageing-related renal fibrosis in male mice by normalizing mitochondrial functions and autophagy. *Biogerontology*, 2022, **23**(6): 731-740
- [118] Shmulevich R, Krizhanovsky V. Cell Senescence, DNA damage, and metabolism. *Antioxid Redox Signal*, 2021, **34**(4): 324-334
- [119] Chien Y, Scuoppo C, Wang X, *et al.* Control of the senescence-associated secretory phenotype by NF-kappaB promotes senescence and enhances chemosensitivity. *Genes Dev*, 2011, **25**(20): 2125-2136
- [120] Alimbetov D, Davis T, Brook A J, *et al.* Suppression of the senescence-associated secretory phenotype (SASP) in human fibroblasts using small molecule inhibitors of p38 MAP kinase and MK2. *Biogerontology*, 2016, **17**(2): 305-315
- [121] Li Q, Ge C, Tan J, *et al.* Juglanin protects against high fat diet-induced renal injury by suppressing inflammation and dyslipidemia *via* regulating NF-kappaB/HDAC3 signaling. *Int Immunopharmacol*, 2021, **95**: 107340
- [122] Bunbupha S, Apaijit K, Maneesai P, *et al.* Nobiletin ameliorates high-fat diet-induced vascular and renal changes by reducing inflammation with modulating AdipoR1 and TGF-beta1 expression in rats. *Life Sci*, 2020, **260**: 118398
- [123] Isaka Y. Targeting TGF-beta signaling in kidney fibrosis. *Int J Mol Sci*, 2018, **19**(9): 2532
- [124] Voelker J, Berg P H, Sheetz M, *et al.* Anti-TGF-beta1 antibody therapy in patients with diabetic nephropathy. *J Am Soc Nephrol*, 2017, **28**(3): 953-962
- [125] Huang W, Hickson L J, Eirin A, *et al.* Cellular senescence: the good, the bad and the unknown. *Nat Rev Nephrol*, 2022, **18**(10): 611-627

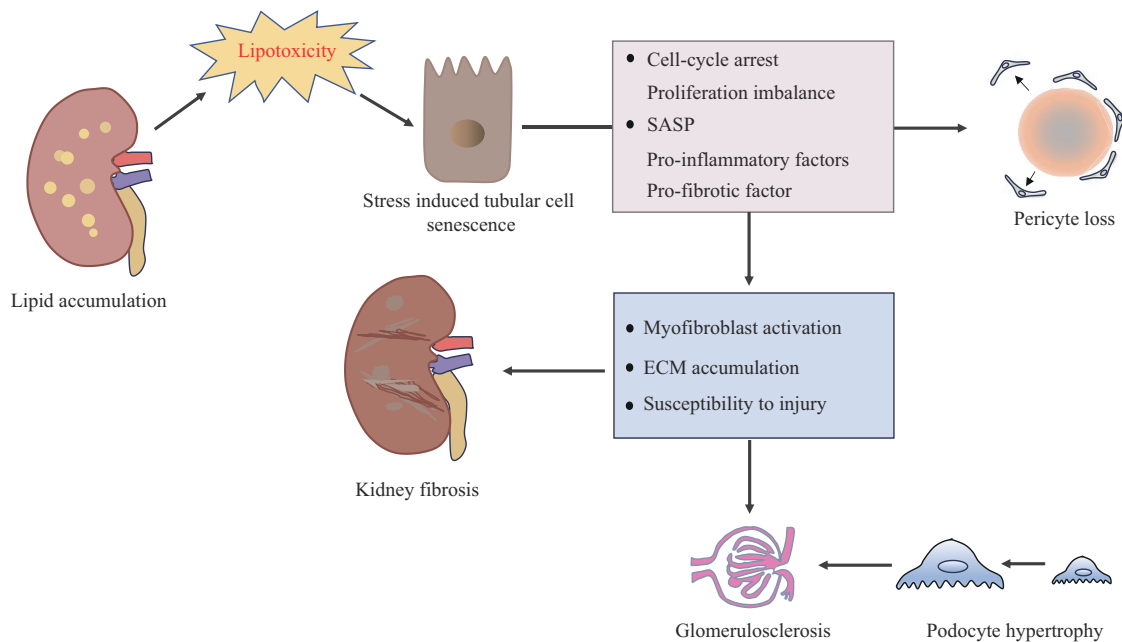
Role of Lipid Metabolism Disorders in Renal Ageing and Renal Fibrosis*

WU Sheng-Quan^{1,2)}, YANG Meng^{1,2)}, LIU Xin-Guang^{1,2)**}

¹⁾Guangdong Provincial Key Laboratory of Medical Molecular Diagnostics, Institute of Aging Research, Guangdong Medical University, Dongguan 523808, China;

²⁾Institute of Biochemistry & Molecular Biology, Guangdong Medical University, Dongguan 523808, China)

Graphical abstract



Abstract Chronic kidney disease (CKD) has become a significant global public health problem. It is defined as chronic renal structural and functional dysfunction caused by various reasons. The prevalence of obesity and diabetes has increased dramatically in developing countries, which substantially affected the patterns of CKD observed in these regions. It's inevitable that the disease spectrum of CKD is converting to metabolic diseases. CKD is also considered an independent risk factor for renal ageing and cardiovascular disease in the elderly, which usually progresses to end-stage renal disease (ESRD). Renal interstitial fibrosis is the pathological basis of ESRD and is a microscopic manifestation of renal ageing. Conversely, renal ageing is a risk factor for interstitial fibrosis. Although the healthy kidney has a relatively low lipid level, CKD-associated dyslipidemia has been extensively

* This work was supported by grants from The National Natural Science Foundation of China (81971329, 82200821) and Guangdong Basic and Applied Basic Research Foundation (2023A1515012838).

** Corresponding author.

Tel: 86-769-22896026, E-mail: liuxg@gdmu.edu.cn

Received: August 3, 2023 Accepted: October 26, 2023

studied. Nevertheless, less is known about the contribution of lipid disorders to the development of renal senescence and interstitial fibrosis. Recent studies have demonstrated that lipid metabolism disorders occur in the progress of renal aging and interstitial fibrosis. Renal lipids accumulate once lipid uptake and synthesis exceed the balance with lipolysis, which is mainly characterized by increased levels of triglyceride (TG) and oxidized low-density lipoprotein, and decreased levels of high-density lipoprotein. Excessive lipid accumulation in the kidney not only induces lipotoxicity and endoplasmic reticulum stress but also increases intracellular and mitochondrial reactive oxygen species, which induce stress injury and senescence in renal tubular epithelial cells. Pro-inflammatory and pro-fibrotic cytokines in a senescence-associated secretory phenotype secreted by senescent renal tubular epithelial cells further accelerate their senescence as well as the occurrence of inflammation and pericyte loss, promoting secretion of extracellular matrix (ECM) and subsequent fibrosis in the tubulointerstitial compartment. In addition, podocyte hypertrophy also leads to glomerulosclerosis. Currently, most of the studies on inhibiting or even reversing renal interstitial fibrosis are still in the experimental stage. What's more, effective drugs to slow down renal aging have not been reported. Many inflammatory and fibrotic factors are both components of the senescence-associated secretory phenotype (SASP), nevertheless, they are not sufficient to recognize cellular senescence. Given that indicators of senescence may vary from disease to disease and organ to organ, there is a need for more sensitive and specific senescence assays. Crucial enzymes and regulatory proteins of lipid metabolic pathways are expected to be potential targets for ameliorating renal aging and interstitial fibrosis. Lipid-lowering approach might represent another therapeutic in the management of kidney injury associated with metabolic dysfunction. Thus, clarifying the molecular regulatory mechanisms of lipid metabolism in kidney is extremely important for the delay of renal aging and the treatment of interstitial fibrosis. This review outlines the effects of lipid metabolism disorders on renal aging and renal fibrosis, analyses the role of lipid metabolism disorders in the development of renal diseases, and summarizes the potential targets and strategies for the prevention of renal aging and renal fibrosis based on lipid metabolism regulation, which will provide a reference for the discovery of new targets for the treatment of renal fibrosis.

Key words renal ageing, renal interstitial fibrosis, lipid metabolism, renal tubular epithelial cells

DOI: 10.16476/j.pibb.2023.0305