

内源性甲醛异常蓄积与记忆衰退*

童志前 韩婵帅 苗君叶 卢静 赫荣乔**

(脑与认知国家重点实验室, 中国科学院生物物理研究所, 北京 10010)

摘要 近期, 本实验室报道了内源性甲醛浓度与认知功能损伤程度之间的关系(*Neurobiol Aging*, 2011, **32**(1): 31-41)。观察到转基因痴呆小鼠(APP、APP/PS1)及衰老加速型(SAMP8)小鼠脑内甲醛浓度较对照组显著升高。参照痴呆鼠脑内甲醛浓度, 实验人员对正常成年小鼠进行注射, 导致其视觉空间记忆能力减退。注射甲醛消除剂可以降低老龄大鼠体内甲醛浓度、减少APP痴呆模型小鼠脑内的老年斑, 且能观察到记忆功能的改善。临床观察显示, 老年痴呆患者尿甲醛浓度与认知功能损伤程度之间呈正相关。甲醛的过量蓄积造成脑慢性损伤, 可能是散发性老年记忆衰退的机制之一。

关键词 痴呆, 甲醛应激, 内源性甲醛, 视觉空间记忆, 记忆衰退

学科分类号 Q5, R74

DOI: 10.3724/SP.J.1206.2011.00241

最近研究发现^[1], 老龄大鼠在记忆力逐步下降的同时, 其海马甲醛含量随老龄化进程而增加, SAMP8小鼠也有类似情况。6月龄APP转基因痴呆小鼠脑甲醛浓度也升高, 并伴有记忆能力的下降, 但12月龄时, 其甲醛浓度与对照相近。APP/PS1转基因痴呆小鼠也有类似的表现。参考痴呆模型鼠脑甲醛浓度, 给正常成年小鼠注射甲醛, 也能观察到其空间记忆能力的下降。这些结果表明, 脑内特别是海马甲醛的蓄积可能是诱发动物体记忆衰退的关键原因之一。

1 内源性甲醛蓄积与记忆衰退

甲醛是地球形成初期出现的有机分子, 存在于生物体内外。人血液中甲醛浓度约为0.06~0.08 mmol/L^[1], 脑组织内的浓度是血液浓度的2~4倍(0.2~0.3 mmol/L)^[2]。Kilburn等^[3]与Perna等^[4]在动物实验及职业性甲醛暴露人员的研究中发现, 气态甲醛暴露可明显引起大鼠记忆能力下降^[5], 并伴有海马和皮层神经元的死亡与丢失^[6-7]。甲醛的吸入能改变大鼠嗅球和海马神经细胞的形态, 海马CA3区CaMK II表达量下降, 也可损害小鼠在水迷宫、六臂放射状水迷宫测试中的能力^[8-11]。

Khokhlov等发现, 伴有记忆衰退的多发性硬化症(multiple sclerosis)患者, 其血液和脑脊液中甲醛含量明显上升, 而给予甘氨酸对病情有缓解作

用^[12-13]。作者研究显示^[1], 正常人血甲醛维持在0.06~0.08 mmol/L ($n=421$), 但随年龄增加有逐渐升高的趋势。根据本实验室调查结果, 不同年龄健康人($n=236$)尿甲醛浓度(约0.06 mmol/L)也有随老龄化进程而增加的趋势。此外, 尿甲醛浓度($n=141$)随痴呆严重程度而增高, 提示尿甲醛浓度与痴呆的发生呈正相关。数据显示, 分别有42%(21/50)轻度痴呆患者, 82.05%中度痴呆患者和88.46%重度痴呆患者尿甲醛浓度高于正常水平(图1)。痴呆伴高血压($n=21$)与单纯性高血压患者相比, 尿甲醛浓度之间具有显著性差异。痴呆伴糖尿病($n=10$)与单纯性糖尿病患者的尿甲醛浓度亦存在显著性差异。尸检结果表明, 痴呆患者($n=8$)海马内甲醛浓度高于正常对照组, 患者皮层甲醛浓度($n=10$)也有增高的趋势。这些结果提示, 老年记忆衰退与内源性甲醛浓度异常升高有关。

* 中国博士后基金(20090460047), 国家重点基础研究发展计划(973)(2010CB912303, 2006CB500703), 国家自然科学基金(30970695)和中国科学院创新方向性项目(CAS-KSCX2-YW-R-119, KSCX2-YW-R-256)资助项目。

** 通讯联系人。

Tel: 010-64889876, Fax: 010-64853625,

E-mail: rongqiaohe@gmail.com

收稿日期: 2011-06-02, 接受日期: 2011-06-13

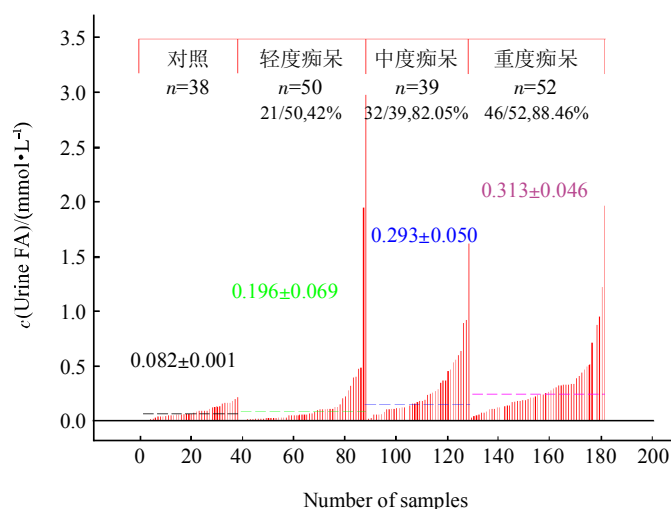


Fig. 1 Urine formaldehyde levels of healthy controls and patients with various degree of dementia

图 1 不同程度老年痴呆患者和健康人尿甲醛水平的比较

2 体内甲醛蓄积的多重机制

2.1 体内甲醛的来源

除环境中的甲醛污染因素外,某些食物、药物(butyric acid, AN-7, mitoxantrone 等)是体内甲醛的又一来源^[14],其余的如汞、甲胺、百草枯等也可以通过线粒体细胞色素 P450 的转化作用产生甲醛^[15].

酶促反应是体内产生甲醛的主要途径。能够催化底物生成甲醛的酶类主要包括 DNA 脱甲基化酶、LSD 脱甲基化酶、胞浆转甲基化酶、内质网脱甲基化酶、氨基脲敏感的胺类氧化酶、脂类氧化酶、线粒体细胞色素 P450 酶等^[15]。其中, DNA 脱甲基化酶能够催化 DNA 脱甲基产生甲醛,这与基因的激活、转录有关^[16-19]。衰老动物^[20]和肿瘤细胞^[21]的整体 DNA 处于低甲基化状态,这也可能是衰老动物和肿瘤细胞内甲醛蓄积的因素之一^[14,22]。

2.2 体内甲醛的清除

甲醛降解酶有依赖谷胱甘肽(GSH)的甲醛脱氢酶(formaldehyde dehydrogenase, FDH or ADH3),非依赖谷胱甘肽的醇类脱氢酶 1 (alcohol dehydrogenases 1, ADH1)和醛脱氢酶 2 (aldehyde dehydrogenases II, ALDH2)。此外, S-甲基谷胱甘肽脱氢酶(S-formylglutathione hydrolase)、醛酮变位酶(glyoxalase II)、过氧化氢酶(catalase)等也能够降解甲醛^[15]。其中, ADH3 在多种组织器官中存在,但其活力因不同组织而有差异^[23], ADH3 在脑蛋白质的表达高于灰质,能抵抗由衰老诱发的神经退

行性变^[24]。研究显示, ADH3 过度表达,可引起果蝇视觉认知功能障碍^[25]。ALDH2 体内分布广泛^[26],其活力异常降低时,可导致记忆能力的下降^[27]。生理条件下,甲醛主要由依赖 GSH 的 ADH3 进行降解;当体内甲醛浓度异常升高时,非依赖 GSH 的 ALDH2 也发挥重要的降解作用^[28]。

ADH3 和 ALDH2 的基因具有多态性,其功能异常与痴呆的发生存在联系^[29-31]。ALDH2 基因敲除小鼠记忆力明显下降^[27,32-33]。APP 被酶切后形成的 amyloid 能抑制线粒体内醇脱氢酶(ADH)的活力,这也可能是甲醛在 APP 转基因模型鼠中蓄积的原因之一。作者实验表明,6 月龄 APP 转基因鼠体内甲醛浓度明显上升^[1]。随着衰老,体内 GSH 含量逐步减少,也可能造成 ADH3 酶活力下降,从而导致机体代谢甲醛的能力降低。

3 甲醛诱发记忆衰退的神经分子机制

甲醛能自发与蛋白质的 α/ϵ 氨基发生反应,导致蛋白质聚积,活性丧失。体外实验表明,低浓度甲醛可以引起神经 Tau 蛋白错误折叠,令其生物学功能丧失,并形成细胞毒性聚积物。甲醛可以透过细胞膜和血脑屏障^[34],引起细胞内 Tau 蛋白的聚积,导致神经细胞变性死亡^[35]。如果甲醛的浓度升高到一定程度,便能与胞内更多蛋白质及其他分子进行反应,造成细胞多方面的损伤。这可能是“甲醛慢性中枢神经系统损伤”的细胞生物学基础^[36-38]。

甲醛也能与蛋白质的半胱氨酸巯基反应^[39-41], 从而封闭蛋白质的侧链巯基^[42-44]. 因此, 甲醛可能与 NMDA 受体 NR2B 亚基上的半胱氨酸巯基反应, 影响 NMDA 受体的功能. 美金刚(Memantine, NMDA 受体的拮抗剂)能与 NMDA 受体 NR2B 亚基的半胱氨酸巯基结合^[45], 从而对老年痴呆产生一

定的疗效. 作者实验表明: 病理浓度的甲醛能够阻断 NMDA 受体, 并抑制海马长时程增强(LTP)的形成, 从而导致大鼠空间学习记忆能力下降. 甲醛浓度异常还可以使皮层编码长时程记忆相关蛋白及其 DNA 甲基化模式被破坏, 诱发海马和皮层神经元死亡, 引起记忆下降(图 2).

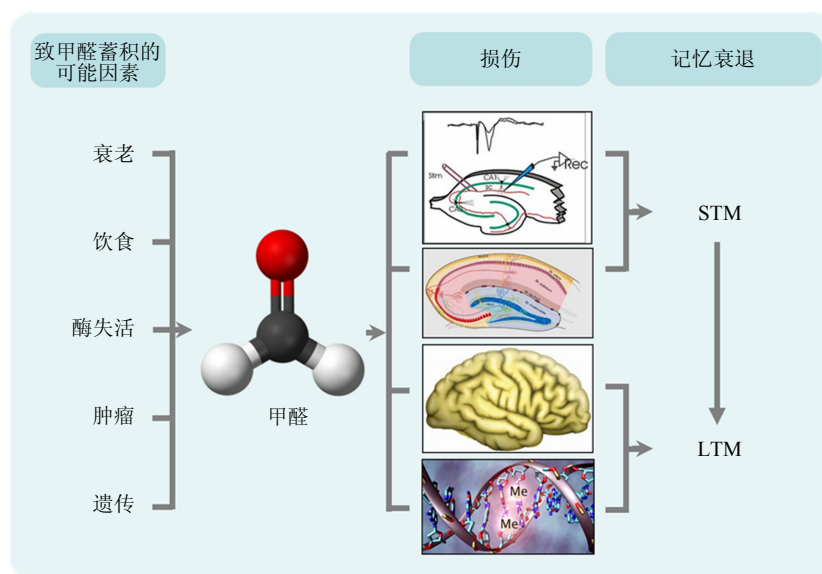


Fig. 2 A putative mechanism of formaldehyde-induced memory decline

图 2 甲醛诱发记忆衰退的可能机制

STM: 短时记忆, LTM: 长时记忆.

4 研究意义和展望

甲醛通过多靶点损伤中枢神经系统, 但其诱发认知功能衰退的机制依然不清楚. 另一方面, 导致体内甲醛蓄积的原因是多方面的, 如环境、药物、肿瘤、老化、基因异常等^[14]. 老年痴呆发生发展的病理机制十分复杂, 病因诸多. 从流行病学的角度来分析, 家族遗传性痴呆仅占老年痴呆病例的 3% 以下, 而散发性痴呆患者的比例则大于 97%. 临床研究表明, 散发性老年痴呆患者伴有尿甲醛浓度显著升高者占所调查全部病例的 30%~40%, 也就是说, 甲醛蓄积造成中枢神经系统慢性损伤, 从而诱发认知功能障碍的病例, 在老年痴呆患者中占有相当的比例. 因此, 作者假设: 甲醛在脑内蓄积造成的脑慢性进行性损伤, 可能是散发性老年痴呆发病的原因之一——^[36,46].

蛋白质寡聚化是其功能调节的一种形式^[47-48]. 但是, 蛋白质变性聚集则是神经系统退行性病变过程中的重要表现, 并且是相关领域的研究重

点^[49-50]. 低浓度甲醛可以引起神经系统重要蛋白质的错误折叠, 并导致蛋白质分子聚集形成神经细胞毒性产物^[51-52]. 一定浓度的白藜芦醇(甲醛清除剂)能降低大鼠体内的甲醛浓度, 对甲醛诱发的记忆衰退有一定治疗效果^[1]. 饮水中添加白藜芦醇能减少 APP 转基因痴呆小鼠脑中的老年斑, 提高其空间记忆能力^[53]. 这些结果提示, 降低内源性甲醛浓度可能成为治疗老年痴呆的新构想.

参 考 文 献

- [1] Tong Z, Zhang J, Luo W, *et al.* Urine formaldehyde level is inversely correlated to mini mental state examination scores in senile dementia. *Neurobiol Aging*, 2011, **32**(1): 31-41
- [2] Heck H D, White E L, Casanova-Schmitz M. Determination of formaldehyde in biological tissues by gas chromatography/mass spectrometry. *Biomed Mass Spectrom*, 1982, **9**(8): 347-353
- [3] Kilburn K H, Warshaw R, Thornton J C. Formaldehyde impairs memory, equilibrium, and dexterity in histology technicians: effects which persist for days after exposure. *Arch Environ Health*, 1987, **42**(2): 117-120
- [4] Perna R B, Bordini E J, Deinzer-Lifrak M. A case of claimed

- persistent neuropsychological sequelae of chronic formaldehyde exposure: clinical, psychometric, and functional findings. *Arch Clin Neuropsychol*, 2001, **16**(1): 33–44
- [5] Malek F A, Moritz K U, Fanghanel J. Formaldehyde inhalation & open field behaviour in rats. *Indian J Med Res*, 2003, **118**: 90–96
- [6] Aslan H, Songur A, Tunc A T, *et al.* Effects of formaldehyde exposure on granule cell number and volume of dentate gyrus: a histopathological and stereological study. *Brain Res*, 2006, **1122**(1): 191–200
- [7] Gurel A, Coskun O, Armutcu F, *et al.* Vitamin E against oxidative damage caused by formaldehyde in frontal cortex and hippocampus: biochemical and histological studies. *J Chem Neuroanat*, 2005, **29**(3): 173–178
- [8] Lu Z, Li CM, Qiao Y, *et al.* Effect of inhaled formaldehyde on learning and memory of mice. *Indoor Air*, 2008, **18**(2): 77–83
- [9] 冯丫娟, 丁书姝, 翟金霞. 维生素 E 拮抗甲醛导致小鼠学习记忆能力改变的作用. *现代预防医学*, 2009, **36**(10): 1833–1835
Feng Y J, Ding S S, Zhai J X. *Modern Preventive Medicine*, 2009, **36**(10): 1833–1835
- [10] 李一乔, 陈浩浩, 尹一飞, 韩飞, 叶学松, 凌树才. 甲醛吸入对大鼠嗅球和海马神经细胞的影响. *浙江大学学报(医学版)*, 2010, **39**(3): 272–278
Li Y Q, Chen H H, Yin Y F, Han F, Ye X S, Ling S C. *J Zhejiang Univ (Medical Sci)*, 2010, **39**(3): 272–278
- [11] 廖双, 蒋莉, 张晓萍. 脑发育不同阶段甲醛暴露对大鼠学习记忆能力及海马 CA3 区 CaMK II 表达的影响. *重庆医科大学学报*, 2010, **35**(3): 342–344
Liao S, Jiang L, Zhang X P. *J Chongqing Univ*, 2010, **35**(3): 342–344
- [12] Khokhlov A P, Zavalishin I A, Savchenko IuN, *et al.* Disorders of formaldehyde metabolism and its metabolic precursors in patients with multiple sclerosis. *Zh Nevropatol Psikhiatr Im S S Korsakova*, 1989, **89**(2): 45–48
- [13] Conaway C C, Whysner J, Verna L K, *et al.* Formaldehyde mechanistic data and risk assessment: endogenous protection from DNA adduct formation. *Pharmacol Ther*, 1996, **71**(1-2): 29–55
- [14] Kalasz H. Biological role of formaldehyde, and cycles related to methylation, demethylation, and formaldehyde production. *Mini Rev Med Chem*, 2003, **3**(3): 175–192
- [15] 童志前, 万有, 罗文鸿, 赫荣乔. 内源性甲醛及其相关人类重大疾病. *自然科学进展*, 2008, **18**(11): 1201–1211
Tong Z Q, Wan Y, Luo W H, He R Q. *Prog Nat & Sci*, 2008, **18**(11): 1201–1211
- [16] Gerken T, Girard C A, Tung Y C, *et al.* The obesity-associated FTO gene encodes a 2-oxoglutarate-dependent nucleic acid demethylase. *Science*, 2007, **318**(5855): 1469–1472
- [17] Roy T W, Bhagwat A S. Kinetic studies of *Escherichia coli* AlkB using a new fluorescence-based assay for DNA demethylation. *Nucleic Acids Res*, 2007, **35**(21): e147
- [18] Treweek S C, Henshaw T F, Hausinger R P, *et al.* Oxidative demethylation by *Escherichia coli* AlkB directly reverts DNA base damage. *Nature*, 2002, **419**(6903): 174–178
- [19] Yi C, Yang C G, He C. A non-heme iron-mediated chemical demethylation in DNA and RNA. *Acc Chem Res*, 2009, **42**(4): 519–529
- [20] Liu L, van Groen T, Kadish I, *et al.* DNA methylation impacts on learning and memory in aging. *Neurobiol Aging*, 2009, **30**(4): 549–560
- [21] Ehrlich M. DNA hypomethylation in cancer cells. *Epigenomics*, 2009, **1**: 239–259
- [22] Thorndike J, Beck W S. Production of formaldehyde from N5-methyltetrahydrofolate by normal and leukemic leukocytes. *Cancer Res*, 1977, **37**(4): 1125–1132
- [23] Uotila L, Koivusalo M. Expression of formaldehyde dehydrogenase and Sformylglutathione hydrolase activities in different rat tissues. *Adv Exp Med Biol*, 1997, **414**: 365–371
- [24] Mori O, Haseba T, Kameyama K, *et al.* Histological distribution of class III alcohol dehydrogenase in human brain. *Brain Res*, 2000, **852**(1): 186–190
- [25] Hou Q, Jiang H, Zhang X, *et al.* Nitric oxide metabolism controlled by formaldehyde dehydrogenase (fdh, homolog of mammalian GSNOR) plays a crucial role in visual pattern memory in *Drosophila*. *Nitric Oxide*, 2011, **24**(1): 17–24
- [26] Mukerjee N, Pietruszko R. Human mitochondrial aldehyde dehydrogenase substrate specificity: comparison of esterase with dehydrogenase reaction. *Arch Biochem Biophys*, 1992, **299**: 23–29
- [27] Ohta S, Ohsawa I. Dysfunction of mitochondria and oxidative stress in the pathogenesis of Alzheimer's disease: on defects in the cytochrome c oxidase complex and aldehyde detoxification. *J Alzheimers Dis*, 2006, **9**(2): 155–166
- [28] Dicker E, Cederbaum AI. Inhibition of the low-Km mitochondrial aldehyde dehydrogenase by diethyl maleate and phorone *in vivo* and *in vitro*. *Biochem J*, 1986, **240**: 821–827
- [29] Li H S, Dai Y F, Huang H L, *et al.* Polymorphisms of aldehyde and alcohol dehydrogenase genes associated with susceptibility to trichloroethylene-induced medicamentosa-like dermatitis. *Wei Sheng Yan Jiu*, 2006, **35**(2): 149–151
- [30] Thomasson H R, Edenberg H J, Crabb D W, *et al.* Alcohol and aldehyde dehydrogenase genotypes and alcoholism in Chinese men. *Am J Hum Genet*, 1991, **48**(4): 677–681
- [31] Deltour L, Foglio M H, Duester G. Metabolic deficiencies in alcohol dehydrogenase Adh1, Adh3, and Adh4 null mutant mice. Overlapping roles of Adh1 and Adh4 in ethanol clearance and metabolism of retinol to retinoic acid. *J Biol Chem*, 1999, **274**(24): 16796–16801
- [32] Nakashima Y, Ohsawa I, Konishi F, *et al.* Preventive effects of Chlorella on cognitive decline in age-dependent dementia model mice. *Neurosci Lett*, 2009, **464**(3): 193–198
- [33] Ohsawa I, Nishimaki K, Murakami Y, *et al.* Age-dependent neurodegeneration accompanying memory loss in transgenic mice defective in mitochondrial aldehyde dehydrogenase 2 activity. *J Neurosci*, 2008, **28**(24): 6239–6249
- [34] Gronvall J L, Garpenstrand H, Orelund L, *et al.* Autoradiographic imaging of formaldehyde adducts in mice: possible relevance for vascular damage in diabetes. *Life Sci*, 1998, **63**(9): 759–768
- [35] Sajjad Haider Naqvi, 王维山, 苗君叶, 赫荣乔. 甲醛诱导 Tau 蛋白形成“孔道样”聚集结构. *生物化学与生物物理进展*, 2010, **37**(11): 1–9
Sajjad Haider Naqvi, Wang W S, Miao J Y, He R Q. *Prog Biochem Biophys*, 2010, **37**(11): 1–9
- [36] 李芳序, 卢静, 许亚杰, 童志前, 聂春来, 赫荣乔. 老年性痴呆发病过程中内源性甲醛慢性损伤机. *生物化学与生物物理进展*,

- 2008, **35**(4): 393–400
- Li F X, Lu J, Xu Y J, Tong Z Q, Nie C L, He R Q. *Prog Biochem Biophys*, 2008, **35**(4): 393–400
- [37] 王维山, 都智慧, 张力, 李文杰, 李沫, 张金玲, 赫荣乔. 正常老年人与阿尔茨海默病患者尿甲醛浓度的研究. *中华老年心脑血管病*, 2010, **12**(8): 721–722
- Wang W S, Du Z H, Zhang L, Li W J, Li M, Zhang J L, He R Q. *Chin J Geriatr Heart Brain Vessel Dis*, 2010, **12**(8): 721–722
- [38] Yuetao Song, Jintang Wang. Overview of Chinese research on senile dementia in mainland China. *Ageing Research Reviews*, 2010: s6–s12
- [39] Kallen R G. Equilibria for the reaction of cysteine and derivatives with formaldehyde and protons. *J Am Chem Soc*, 1971, **93**(23): 6227–6235
- [40] Kallen R G. The mechanism of reactions involving Schiff base intermediates. Thiazolidine formation from L-cysteine and formaldehyde. *J Am Chem Soc*, 1971, **93**(23): 6236–6248
- [41] Mackenzie C G, Harris J. N-formylcysteine synthesis in mitochondria from formaldehyde and L-cysteine via thiazolidinecarboxylic acid. *J Biol Chem*, 1957, **227**(1): 393–406
- [42] Metz B, Kersten G F, Baart G J, *et al.* Identification of formaldehyde-induced modifications in proteins: reactions with insulin. *Bioconjug Chem*, 2006, **17**(3): 815–822
- [43] Metz B, Kersten G F, Hoogerhout P, *et al.* Identification of formaldehyde-induced modifications in proteins: reactions with model peptides. *J Biol Chem*, 2004, **279**(8): 6235–6243
- [44] Toews J, Rogalski J C, Clark T J, *et al.* Mass spectrometric identification of formaldehyde-induced peptide modifications under *in vivo* protein cross-linking conditions. *Anal Chim Acta*, 2008, **618**(2): 168–183
- [45] Stuart A. Lipton. Pathologically activated therapeutics for neuroprotection. *Nature Reviews Neuroscience*, 2007, **8**: 803–808
- [46] He R Q, Jing L, Miao J Y. Formaldehyde stress. *Science China, Life Sciences*, 2010, **53**(12): 1399–1404
- [47] Feng Y J, Zhang M, Hu M X, *et al.* Disassembly intermediates of RbsD protein remain oligomeric despite the loss of an intact secondary structure. *Sci China Ser C-Life Sci*, 2009, **52**: 997–1002
- [48] Shi Y G. Assembly and structure of protein phosphatase 2A. *Sci China Ser C-Life Sci*, 2009, **52**: 135–146
- [49] Hu J, Li G, Wang H, *et al.* Prevention of pathological change and cognitive degeneration of Tg2576 mice by inoculating Abeta(1-15) vaccine. *Sci China Ser C-Life Sci*, 2008, **51**: 743–750
- [50] Zheng Y P, He J S, Hong T. Biomarkers of Alzheimer's disease in body fluids. *Sci China Ser C-Life Sci*, 2009, **52**: 490–496
- [51] Nie C L, Wei Y, Chen X Y, *et al.* Formaldehyde at low concentration induces protein Tau into globular amyloid-like aggregates *in vitro* and *in vivo*. *PLoS ONE*, 2007(2): e629
- [52] Nie C L, Wang X S, Liu Y, *et al.* Amyloid-like aggregates of neuronal tau induced by formaldehyde promote apoptosis of neuronal cells. *BMC Neurosci*, 2007(8): 9
- [53] Karuppagounder S S, Pinto J T, Xu H, *et al.* Dietary supplementation with resveratrol reduces plaque pathology in a transgenic model of Alzheimer's disease. *Neurochem Int*, 2009, **54**(2): 111–118

Excess Endogenous Formaldehyde Induces Memory Decline*

TONG Zhi-Qian, HAN Chan-Shuai, MIAO Jun-Ye, LU Jing, HE Rong-Qiao**

(State Key Laboratory of Brain and Cognitive Sciences, Institute of Biophysics, The Chinese Academy of Sciences, Beijing 100101, China)

Abstract This review focuses on the relationship between excess endogenous formaldehyde and memory decline, as well as multi-factors inducing formaldehyde abnormal accumulation and cognitive impairments. Elevation of endogenous formaldehyde concentrations in Alzheimer's animal models and clinical patients was discussed. Injection with formaldehyde (referred to the detected level in the AD animal models) into normal mice obviously induced spatial memory decline. Aging, and some of genetic factors, diet and environmental pollutants led to formaldehyde abnormal accumulation. Formaldehyde scavengers could rescue spatial memory of APP-transgenic mice. These studies addressed that excess formaldehyde-induced damage of brain is one of the critical factors of cognitive impairments of sporadic senile dementia. Scavenging excess formaldehyde may be a novel therapy for Alzheimer's disease.

Key words senile dementia, formaldehyde stress, endogenous formaldehyde, visual spatial memory, memory decline

DOI: 10.3724/SP.J.1206.2011.00241

*This work was supported by grants from Chinese Postdoctoral Fund 20090460047, National Basic Research Program of China (2010CB912303; 2006CB500703), The National Natural Science Foundation of China (30970695), and Innovative Project of Chinese Academy of Sciences (CAS-KSCX2-YW-R-119, KSCX2-YW-R-256).

**Corresponding author.

Tel: 86-10-64889876, Fax: 86-10-64853625, E-mail: rongqiaohe@gmail.com

Received: June 2, 2011 Accepted: June 13, 2011